

Curriculum Vitae



Name: Mr. Pathamet Khositharattanakool

Date of birth: November 23th, 1981

Place of birth: Chiang Mai, Thailand

Work office: School of Medicine, Mae Fah Luang University
333 Moo 1, Thasud Subdistrict, Muang District,
Chiang Rai 57100, Thailand

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Education:

2013 Ph.D. (Parasitology)
Department of Parasitology,
Faculty of Medicine,
Chiang Mai University, Chiang Mai, Thailand (GPA: 4.00)

2002 B.Sc. (Medical Technology), Second Class Honor
Faculty of Associated Medical Sciences,
Chiang Mai University, Chiang Mai, Thailand (GPA: 3.47)

Fields of study: Molecular Parasitology
Medical Parasitology

Publications:

1. Jariyapan N, Uparanukraw P, Wannasarn A, Saeung A, **Khositharattanakool P**, Sor-suwan S, Phattanawiboon B. Analysis of haemolymph proteins in the *Brugiamalayi*-susceptible mosquito, *Aedestogoi*, using SDS-PAGE and two-dimensional gel electrophoresis. International Journal of Parasitology Research 2011; 3(2): 31-8.
2. **Khositharattanakool P**, Morakote N, Uparanukraw P. The vasculature of nurse cells infected with non-encapsulated *Trichinella* species. The Southeast Asian Journal of Tropical Medicine and Public Health 2013; 44(4): 561-7.
3. **Khositharattanakool P**, Morakote N, Siriaunkgul S, Uparanukraw P. Expression of vascular endothelial growth factor during nurse cell formation in *Trichinella spiralis* and *Trichinella pseudospiralis* infections. International Journal of Parasitology Research 2013; 5(1): 111-5.

Presentations:

1. **Khositharattanakool P**, Kongkerd N, Uparanukraw P. Cloning and characterization of a gene encoding serine protease inhibitor from *Gnathostomaspingerum*. The Joint International Tropical Medicine Meeting. November 30-December 2, 2005 in Bangkok, Thailand.
2. **Khositharattanakool P**, Uparanukraw P, Appleton JA. Expression levels of *Trichinella spiralis* genes during different developmental stages. RGJ-Ph.D. Congress XI. April 1-3, 2010 in Chonburi, Thailand.
3. **Khositharattanakool P**, Uparanukraw P, Appleton JA. Expression levels of *Trichinella spiralis* genes during different developmental stages. RGJ Seminar Series LXXIV. September 3, 2010 in Chiang Mai, Thailand.

Laboratory techniques and skills:

1. Genomic DNA and RNA extraction
2. *In vivo* and *in vitro* culture of *Trichinella* sp.
3. PCR, RT-PCR, Nested-PCR, Multiplex-PCR, Real time PCR and PCR-RFLP
4. Cell and tissue culture

5. Cytotoxicity test
6. *In vitro* transcription
7. RNA interference technique
8. cDNA library construction
9. Gene cloning
10. Viral transfection
11. Bacterial transformation
12. Bacterial protein expression
13. Protein extraction and purification
14. Phage display screening technique
15. DNA sequencing
16. SDS-PAGE
17. Hybridization; Southern blot, Northern blot, Western blot, colony/plaque lift and *in situ* hybridization
18. ELISA
19. Frozen/paraffin-embedded tissue preparation
20. Immunohistochemistry
21. Bioinformatics
22. Animal research
23. Microinjection of parasitic nematode using micromanipulator

Training experience:

1. HyperCourse on Bioinformatics at Faculty of Science, Mahidol University, Bangkok, Thailand, 2004.
2. Training on collection of excretory-secretory product of *Trichinella spiralis* L1, Vhh phage display screening and *Trichinella spiralis* newborn larva infection by mouse injection at Baker Institute for Animal Health, College of Veterinary Medicine, Cornell University, Ithaca, New York, United States of America, 2008.
3. Training on green fluorescent protein plasmid injection in *Trichinella spiralis* L1 gonad using micromanipulator at University of Pennsylvania, Philadelphia, Pennsylvania, United States of America, 2008.

4. Attendance in The eighth annual retreat of program in infection and pathology, Cornell University at Ramada Geneva Lakefront, Geneva, New York, United States of America, 2008.
5. Certificate in the care and use of animals for students and staff, certified by AALAS Learning Library. Animal Care and Use in Research and Education, United States of America, 2008.
6. Workshop on basic mouse techniques at College of Veterinary Medicine, Cornell University, Ithaca, New York, United States of America, 2008.
7. Workshop on NeonTM Transfection System; New tool for RNAi workflow at Gibthai Training Center, Chiang Mai, Thailand, 2009.
8. Workshop on Fast & simple 2D electrophoresis for complex protein analysis at Gibthai Training Center, Chiang Mai, Thailand, 2009.
9. Lecture attendance on Geographic Information System in Health Science by Prof. Kimberly N. Irvine (Buffalo State University) at Department of Parasitology, Faculty of Medicine, Chiang Mai University, Chiang Mai, Thailand, 2009.
10. Medical Writing Workshop at Alpine golf resort, Chiang Mai, Thailand, 2010.
11. Lecture attendance on more tips for reviewers to like, and editors to publish, your manuscripts in competitive scientific journals by Prof. Bruce G. Weniger at Faculty of Medicine, Chiang Mai University, Chiang Mai, Thailand, 2013.

Teaching experience:

1. Teaching assistance under supervision of Prof. Dr. Judy Appleton in Western blot analysis for undergraduate students of Cornell University during March-October 2008 at Baker Institute for Animal Health, College of Veterinary Medicine, Cornell University.
2. Teaching assistance under supervision of Assoc. Prof. Dr. Pichart Uparanukraw in conventional PCR technique for graduate students during 2005-2012 at Department of Parasitology, Faculty of Medicine, Chiang Mai University.